

(Roca, W., L. Mroginski, L., eds.). Cali, Colombia.

Krikorian A. H., Irizarry, S., Cronauer-Mitra & E. Rivera. 1993. Clonal fidelity and variation in plantain (*Musa* AAB) regenerates from vegetative stem and floral axis tips *in vitro*. *Annals of Botany* 71:519-535.

Lebot V., K. Aradhya, R. Manshardt & B. Meilleur. 1993. Genetic relationships among cultivated bananas and plantains from Asia and the Pacific. *Euphytica* 67: 163-175.

Lee M. 1988. The chromosomal basis of somaclonal variation. *Annual Review of Plant Molecular Biology* 39:413-437.

Lindsay K. & M. Jones. 1989. Biotecnología vegetal agrícola. Editorial Acribia. S.A. Zaragoza. España.

Pérez L. 1992. Comparación de varios métodos de propagación en banano. *CORBANA* 16 (38): 28-32

Reuveni O. 1989. Methods for detecting somaclonal variants in "Williams" bananas. Pp. 316-337 *in* Sigatoka leaf spot diseases. (Fullerton, R., R. Stover, eds).

Reyes L.M., O. Martínez & M. Beltrán. 1997. Quimiovariabilidad en el género *Musa*: II. Caracterización genética mediante nueve sistemas enzimáticos. (submitted).

Sokal R. & Y. Sneath. 1973. Numerical Taxonomy. W.H. Freeman and company. San Francisco. 573 pp.

Simmonds N. & K. Shepherd. 1955. Taxonomy and origins of cultivated bananas. *Journal of Linnean Society of London Botany* 55:302-312.

Simmonds N., K. Shepherd & T. Weatherup. 1990. Numerical taxonomy of the wild bananas (*Musa*). *New Phytologist* 115:567-571.

Teisson C. & E. De Langhe. 1989. Biotechnologies for banana and plantain. Pp. 241-243 *in* Plant biotechnologies for developing countries. Ed Ebenezer Baylis. The Trinity Press, UK.

Ventura J., M. Rojas, E. Yera, J. Lopez & A. Rodriguez. 1987. Variación somaclonal en material de plátano (*Musa* spp.) obtenido mediante micropropagación *in vitro*. *Ciencia Tecnología Agrícola, Viandas Tropicales* 11(1):7-16. Cuba.

Vuytsteke C., R. Swennen, G. Wilson & E. De Langhe. 1988. Phenotypic variation among *in vitro* propagated plantain (*Musa* sp. cultivar "AAB"). *Scientia Horticulturae* 36: 79-88.

Orlando Martínez W. and **Luz Marina Reyes C.** are respectively Professor and Associate Professor at *Facultad de Agronomía, Universidad Nacional de Colombia*. A.A. 14490, Bogota, Colombia.

Margarita Beltrán is a biologist at *Universidad de los Andes*, Bogotá, Colombia.

from suckers and bit material are being compared in replicated plots.

Plant symptoms

Chlorotic and necrotic streaking have been commonplace but the symptoms have been periodic in expression and virus concentration in the leaves has widely fluctuated. A range of unusual symptoms have been noted on some of the infected plants which include the following:

- Leaf bases falling away on pseudostem
- Narrow thicker leaves
- General leaf distortion
- Choking
- Broad yellow lines in the leaf blade parallel to the midrib
- Purple margin on leaf blade
- Leaf twisting
- Abnormal arrangement of leaves on pseudostem (Traveller's Palm appearance)
- Grooves in bases of petioles

The first four symptoms have been described before by Lassoudière (1979), the others are new. Some of the symptoms listed above are usually more dramatic than the chlorotic and necrotic streaking but are not diagnostic, as they may also be caused by stress from other sources. However, as these symptoms may be suggestive of infection with BSV they can be helpful in the detection of likely infected plants, which then need to be confirmed by the presence of the more typical streak symptoms, or laboratory assays.

Minimal effects of BSV on plant crop yields

Results from the plant crop (Table 1) show that BSV had a minor effect on yield. The only yield parameter that was significantly affected by infection with BSV was time from planting to harvest, which was delayed by 3 weeks in infected plants, contributing to a 7% reduction in yield per unit of time. No other measurements of yield were significantly affected by the infection. We are continuing to examine growth of the first ratoon crop.

Implications

All plants in the trial are being tested regularly for virus, and thus far, there is no evidence of spread of BSV into the healthy treatment. At this early stage of the investigations it would appear that the current impact of this strain of BSV in commercial plantings of cv. Williams ((5% infection) is likely to be very minor. Environmental conditions in north Queensland (temperature, soil moisture and soil fertility) are close to optimal, so the plant is

Diseases

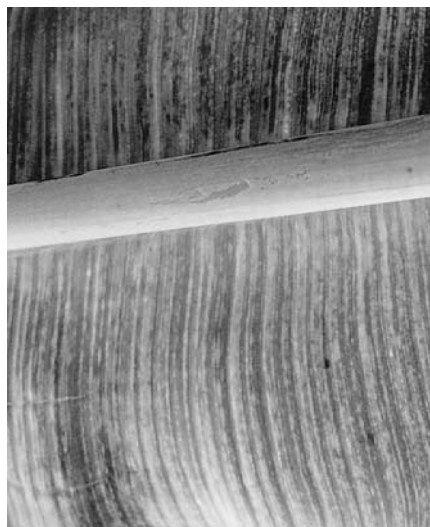
BSV in Australia

Banana streak virus investigations in Australia

Jeff Daniells, Andrew Geering and John Thomas

Banana streak virus (BSV) was first detected in Australia in 1992 in the banana variety Mysore. It has now been found in a range of varieties in Queensland and New South Wales. There have been significant outbreaks on two commercial Cavendish (cv. Williams) plantings at Innisfail. About 5% of plants in these plantations are infected. The properties have been quarantined to prevent further dissemination of infected planting material.

Relatively little is known about BSV compared with some of the other banana infecting viruses and knowledge of the effects of the virus on yield is scanty. The only notable study was



Chlorotic streaking of cv. Williams.

done in the Ivory Coast (Lassoudière 1979), and it was found that BSV caused a yield reduction of Cavendish (cv. Poyo) bananas of between 7 and 90%. One of the major objectives of our project on BSV has therefore been to investigate the effects of an Australian strain of BSV on yield of Cavendish (cv. Williams) bananas under local environmental conditions. A field trial was initiated at South Johnstone Research Station in October 1996 in which yields of BSV-infected plants and indexed BSV-free plants established

Table 1. Effect of BSV infection on plant development, yield and fruit quality.

Treatment	Planting to Harvest (days)	Bunch Wt (kg)	Bunch Wt/Year (kg/plant/year)	% Extra Large Fruit ^α	Hand 3 Finger Length (cm)	Pseudostem Height (cm)	Pseudostem Girth (cm)
BSV -Infected	376 ⁺	22.3	21.6	58.2	24.4	203	48.4
BSV - Free	355	22.7	23.3	62.2	24.6	205	49.1
F Test	*	NS **	*	NS	NS	NS	NS

⁺ Values shown are the means of 12 replicated plots (randomized complete block design - 10 sample plants/plot)

^{*} Significantly different at $P \geq 0.05$

^{**} Differences not significant

^α Extra Large Fruit ≥ 23.5 cm in length

probably able to compensate for any damage caused by infection. However, in more marginal environments, where plants are under greater overall stress, the effects of infection may be greater. The reaction of Cavendish bananas to other BSV strains (Lockhart and Olszewski 1993) may also differ.

In view of experience from overseas (Frans Weilemaker pers. comm.) where BSV effects have increased over time it would seem prudent to maintain quarantine of infected plantings, seek progressive eradication of infected blocks and especially ensure that safeguards on *in vitro* generated planting materials are maintained.

Acknowledgments

These BSV investigations are being funded by the Queensland Banana Industry Protection Board and the Horticulture Research and Development Corporation. ■

References

Lassoudière, A. 1979. Mise en évidence des répercussions économiques de la mosaïque en tirets du bananier en Côte d'Ivoire. Possibilités de lutte par éradication. *Fruits* 34(1): 3-34.

Lockhart, B.E.L. & Olszewski, N.E. 1993. Serological and genomic heterogeneity of banana streak badnavirus: implications for virus detection in *Musa* germplasm. Pp. 105-113 in *Breeding Banana and Plantain for Resistance to Diseases and Pests*. (Ganry J., ed.) CIRAD/INIBAP, Montpellier, France.

Jeff Daniells is horticulturist at Queensland Horticulture Institute, Department of Primary Industries, Queensland, PO Box 20, South Johnstone QLD 4859, Australia. **Andrew Geering** and **John Thomas** work also at QDPI at the following address: 80 Meiers Road, Indooroopilly, QLD 4068, Australia.



Distorted leaves.



Purple leaf margins.



Choking of diseased plant.



Traveller's palm appearance of diseased plant.



Leaf bases falling away from pseudostem.



Broad yellow lines in the leaf blade.



Groove in petiole base.